

# SmartCells™



A Division of Gene Therapy Systems, Inc.

## High Efficiency Chemically Competent *E. coli*

Cat. #	Contents	Quantity
C101020	SmartCells™ Chemically Competent <i>E. coli</i> (>1 x 10 <sup>9</sup> cfu/μg)	20 x 50 μl
	SOC Medium	6 ml
	pUC19 Positive Control Plasmid	20 μl (10pg/μl)
C101120	SmartCells™ F' Chemically Competent <i>E. coli</i> (>1 x 10 <sup>9</sup> cfu/μg)	20 x 50 μl
	SOC Medium	6 ml
	pUC19 Positive Control Plasmid	20 μl (10pg/μl)

<b>Shipping</b>	Shipped on Dry Ice
<b>Storage</b>	Store cells at -70°C; store the SOC Medium at room temperature. Stable for 6 months.

Related Products	Catalog #
EZ-Spread™ Beads, Single-Use Tubes, 50 tubes	C400050
EZ-Spread™ Beads, Dispenser Bottle	C400100

**Introduction:** Most commercially available chemically competent cells, regardless of claimed efficiency, frequently underperform in real transformation experiments, especially when using ligation mixtures instead of supercoiled DNA. SmartCells™ chemically competent *E. coli* are prepared by a unique procedure to warrant the highest and most robust transformation performance under diverse conditions. There is no need to dilute or purify your ligation mix before transformation. If needed, over 10 μl of full strength ligation mix can be added to 50 μl competent cells without significantly compromising transformation results. SmartCells™ carry genotypes suitable for most cloning needs such as blue/white selection, generation of plasmid vector based libraries or gene banks, and ability to be transformed efficiently with large plasmids. SmartCells™ F' is also available for use with M13 cloning vectors.

SmartCells™ F <sup>-</sup> : <i>recA1 endA1 hsdR17 supE44 thi-1 gyrA96 relA1 φ80lacZΔM15 Δ(lacZYA-argF)U169</i>
SmartCells™ F <sup>'</sup> : <i>recA1 endA1 hsdR17 supE44 thi-1 gyrA96 relA1 φ80lacZΔM15 Δ(lacZYA-argF)U169</i>

## METHODS AND PROCEDURES

### A. Transformation Protocol

1. Thaw one tube of the SmartCells™ competent cells on ice (10-15 minutes).
2. Add 1-10 μl of ligation mix to the cells; mix gently and incubate on ice for 15 to 30 minutes.
3. Heat the mix at 42°C for 45 seconds.
4. Add 0.25 ml room temperature SOC medium and incubate at 37°C for 1 hour in an air incubator. Shaking tubes horizontally at 225 rpm is recommended for the best efficiency.
5. Dilute transformation reaction if necessary and spread 100 μl of transformed cells on LB/Agar plates containing appropriate selection (e.g. ampicillin or kanamycin). Alternatively, if maximum numbers of colonies are desired, collect cells by spinning in a microfuge for 10 seconds. Resuspend cell pellet in 50 μl SOC and spread on the agar plates containing antibiotics.
6. Incubate overnight at 37°C.

### B. Notes

- It is not necessary to dilute your ligation mix with TE. In our test, up to 10 μl of undiluted ligation mix could be used without significantly compromising the transformation efficiency.
- Transformation efficiencies for ligation of inserts to vectors will be 2 - 5 fold lower than obtained with supercoiled plasmids.
- If unexpectedly lower number of colonies are observed, we recommend to test the efficiency of competent cells using the provided supercoiled pUC19 plasmid DNA as follows:
  1. Transform 5 μl (50 pg) pUC19 into 50 μl of competent cells.
  2. Repeat steps 2-6.
  3. Dilute the transformation reaction 50 fold with SOC and plate 30 μl on a LB agar plate containing 100 μg/ml ampicillin.
  4. Incubate overnight at 37°C and count colonies. Calculate transformation efficiency as follows:

$$\frac{\text{Number of Colonies}}{50 \text{ pg pUC19}} \times \frac{1 \times 10^6 \text{ pg}}{\mu\text{g}} \times \frac{300 \mu\text{l}}{30 \mu\text{l plated}} \times 50^\dagger = \text{CFU}/\mu\text{g}$$

† 50 is the dilution factor

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